



King's Research Portal

DOI:

[10.1111/all.12939](https://doi.org/10.1111/all.12939)

Document Version

Peer reviewed version

[Link to publication record in King's Research Portal](#)

Citation for published version (APA):

Roberts, G., Ollert, M., Aalberse, R., Austin, M., Custovic, A., DunnGalvin, A., Eigenmann, P. A., Fassio, F., Grattan, C., Hellings, P., Hourihane, J., Knol, E., Muraro, A., Papadopoulos, N., Santos, A. F., Schnadt, S., & Tzeli, K. (2016). A new framework for the interpretation of IgE sensitization tests. *Allergy*, 71(11), 1540-1551. <https://doi.org/10.1111/all.12939>

Citing this paper

Please note that where the full-text provided on King's Research Portal is the Author Accepted Manuscript or Post-Print version this may differ from the final Published version. If citing, it is advised that you check and use the publisher's definitive version for pagination, volume/issue, and date of publication details. And where the final published version is provided on the Research Portal, if citing you are again advised to check the publisher's website for any subsequent corrections.

General rights

Copyright and moral rights for the publications made accessible in the Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognize and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the Research Portal

Take down policy

If you believe that this document breaches copyright please contact librarypure@kcl.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.

Received Date : 23-Mar-2016

Revised Date : 16-May-2016

Accepted Date : 23-May-2016

Article type : Position Paper

A new framework for the interpretation of IgE sensitization tests

Authorship

Graham Roberts¹, Markus Ollert², Rob Aalberse³, Moira Austin⁴, Adnan Custovic⁵, Audrey DunnGalvin⁶, Philippe A Eigenmann⁷, Filippo Fassio⁸, Clive Grattan⁹, Peter Hellings¹⁰, Jonathan Hourihane¹¹, Edward Knol¹², Antonella Muraro¹³, Nikolaos Papadopoulos^{14,15}, Alexandra F. Santos¹⁶⁻¹⁸, Sabine Schnadt¹⁹, Kassiani Tzeli¹⁵.

Affiliations

1. David Hide Asthma and Allergy Research Centre, St Mary's Hospital, Isle of Wight; NIHR Respiratory Biomedical Research Unit, University Hospital Southampton NHS Foundation Trust and Clinical and Experimental Sciences Academic Unit, University of Southampton Faculty of Medicine, Southampton, United Kingdom
2. Klinik und Poliklinik für Dermatologie und Allergologie, Klinikum rechts der Isar and Technische Universität München, Biedersteiner Str. 29, D-80802 München, Germany
3. Sanquin Research, Dept of Immunopathology, Amsterdam and Landsteiner Laboratory, Academic Medical Center, University of Amsterdam, Amsterdam, The Netherlands

This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the Version of Record. Please cite this article as doi: 10.1111/all.12939

This article is protected by copyright. All rights reserved.

- Accepted Article
4. Anaphylaxis Campaign, United Kingdom
 5. Institute of Inflammation and Repair. University of Manchester, University Hospital of South Manchester, Manchester, United Kingdom
 6. School of Applied Psychology; Department of Paediatrics and Child Health; University College Cork, Ireland
 7. Department of Child and Adolescent, University Hospitals of Geneva, Geneva Switzerland
 8. Careggi Hospital, Florence, Italy
 9. Dermatology Centre, Norfolk & Norwich University Hospital, Norwich, United Kingdom
 10. University Hospitals Leuven, Leuven, Belgium
 11. Paediatrics and Child Health, University College Cork, Ireland
 12. Departments of Immunology and Dermatology/Allergology, University Medical Center Utrecht. Utrecht, The Netherlands
 13. Department of Pediatrics, University of Padua, Padova, Italy
 14. Centre for Pediatrics and Child Health, Institute of Human Development, University of Manchester, United Kingdom
 15. Allergy Dpt, 2nd Pediatric Clinic, University of Athens, Greece
 16. Department of Paediatric Allergy, Division of Asthma, Allergy & Lung Biology, King's College London, United Kingdom
 17. MRC & Asthma UK Centre in Allergic Mechanisms of Asthma, London, United Kingdom
 18. Immunoallergology Department, Coimbra University Hospital, Coimbra, Portugal

19. German Allergy and Asthma Association (Deutscher Allergie- und Asthmabund (DAAB)), Mönchengladbach, Germany

Correspondence

Professor Graham Roberts

University of Southampton Faculty of Medicine (MP803), Southampton General Hospital,
Tremona Road, Southampton SO16 6YD, United Kingdom

Tel.: (023) 8120 6160

Fax: (023) 8087 8847

E-mail: g.c.roberts@soton.ac.uk

Key words: IgE sensitization tests, clinical allergy, skin prick testing, serum specific IgE

Abstract (250 words)

IgE sensitization tests, such as skin prick testing and serum specific IgE, have been used to diagnose IgE-mediated clinical allergy for many years. Their prime drawback is that they detect sensitization which is only loosely related to clinical allergy. Many patients therefore require provocation tests to make a definitive diagnosis; these are often expensive and potentially associated with severe reactions. The likelihood of clinical allergy can be semi-quantified from an IgE sensitization test results. This relationship varies though according to the patients' age, ethnicity, nature of the putative allergic reaction and co-existing clinical diseases such as eczema. The likelihood of clinical allergy can be more precisely estimated

from an IgE sensitization test result, by taking into account the patient's presenting features (pre-test probability). The presence of each of these patient specific factors may mean that a patient is more or less likely to have clinically allergy with a given test result (post-test probability). We present two approaches to including pre-test probabilities in the interpretation of results. These approaches are currently limited by a lack of data to allow us to derive pre-test probabilities for diverse setting, regions and allergens. Also, co-factors, such as exercise, may be necessary for exposure to an allergen to result in an allergic reaction in specific IgE positive patients. The diagnosis of IgE-mediated allergy is now being aided by the introduction of allergen component testing which may identify clinically relevant sensitization. Other approaches are in development with basophil activation testing being closest to clinical application.

Introduction

Allergies are the most frequent chronic diseases in Europe today, affecting the daily lives of more than 60 million people (1). Typical manifestations are asthma, rhinitis, eczema, food allergy and anaphylaxis. Many non-allergic disorders present with symptoms that are similar to allergic diseases making it important to have good tests for allergy. There are also many situations where a patient may be exposed to more than one putative allergen, so tests to identify the triggering allergen are required. Allergic diseases can be divided into IgE mediated and non-IgE mediated allergy (2). IgE mediated allergy, with rapid onset of symptoms after exposure to the allergen, is associated with specific IgE to the relevant allergen. This report will focus on IgE-mediated allergy as we have a number of routine clinical tests that can detect the presence of specific IgE. Skin prick testing (SPT) can provide immediate information about the presence of IgE sensitization to specific allergens in clinic (3). Serum specific IgE testing requires blood to be sent to a laboratory and can be tested to allergen extracts or individual allergen components (4). Atopy patch testing has

Accepted Article

been developed over the last decade to understand the relationship between IgE sensitization and the cell-mediated response that characterizes atopic eczema but is not widely used (5). Other approaches to testing for IgE-mediated allergy, such as basophil activation test, have been used in the research setting and their application for clinical use is in development (6). The drawback of IgE sensitization tests is that they report the presence of specific IgE, not the presence of clinical allergy (Figure 1). However, when used by clinicians trained in their interpretation, they are useful to support a diagnosis of clinical allergy and identify the likely triggering allergens within the context of a full clinical history.

There have been considerable advances in allergy over the last decade. Firstly, we now have a better understanding of the pathophysiology of allergic disease and immunological impact of therapies. Secondly, technological advances have allowed us to detect individual allergen components. Finally, with better datasets, we are beginning to understand the relationship between specific presentations, IgE sensitization test results and clinical allergy. For these reasons, the European Academy of Allergy and Clinical Immunology (EAACI) formed a Task Force to reassess how we employ IgE sensitization tests. The Task Force involved a wide range of clinicians, scientists and patient groups. The aim of the Task Force was to refine our conceptual approach to interpreting IgE test results by integrating information from the patient history with the test results and to consider additional tests to improve our ability to accurately diagnose clinical allergy to allow us to improve patient care. Although many of the examples used are from food allergy, given that we have much better evidence in this area, the approach is a generic one that should be applicable to any allergic disease. This position paper builds on the activities of other EAACI diagnostic task forces including those focused on children (7), rhinitis (8) and food allergy (9, 10). While this position paper is primarily aimed at allergists, the Task Force is planning to disseminate the concepts more widely using focused materials aimed at different audiences including specialty physicians, primary care doctors and patients.

The relevance of IgE sensitization to clinical allergy

Pathophysiology of IgE-mediated allergy

An understanding of the pathophysiology of IgE-mediated allergy is useful for considering how to best interpret allergy test results. Mast cell degranulation commences with cross-linking of IgE receptors by allergen, functional autoantibodies against IgE or the high affinity IgE receptor (FcεRI), or FcεRI-independent such as opiates or C5a complement. This results in calcium-dependent series of intracellular signalling events causing a piecemeal or anaphylactic degranulation of tissue mast cells and circulating basophils. Preformed mediators, such as histamine and the generation of vasoactive eicosanoids, induce an acute cellular inflammatory response. Longterm effects on inflammation are influenced by local tissue factors and structures.

The immediate effects of histamine release depend on the organ affected (Box 1). They include itching, redness, weals and angio-oedema of the skin; wheezing of the lung; rhinorrhoea, nasal blockage and sneezing; lacrimation and itching of the eye; vomiting, diarrhoea and abdominal pain of the bowel and faintness and tachycardia of the circulation, culminating in collapse if the reaction is severe. Understanding how organs are affected differently by the same downstream stimulus-secretion events in the mast cell and basophil resulting in allergic disease and how IgE-induced hypersensitivity may be abrogated by the development of immunological tolerance is central to understanding allergy.

Development of tolerance in IgE-mediated allergy

The development of natural tolerance is complex and poorly understood but the events involved in specific allergen immunotherapy have been closely studied (11). They involve the induction of peripheral T-cell tolerance by promotion of regulatory Treg and Tr1 cells. These directly or indirectly suppress pro-inflammatory cells including eosinophils, mast cells and basophils by formation of specific IgG₄, followed by reduction in specific IgE.

Diagnosis of IgE sensitization in clinical practice

The spectrum of clinical disease linked with IgE-sensitisation includes anaphylaxis, asthma, atopic dermatitis, conjunctivitis, drug reactions, eosinophilic enteritis, food allergies, oral allergy syndrome, rhinitis, urticaria and venom allergy (examples in Box 1). The primary tool for assessing immediate hypersensitivity reactions is the history. Investigations are, to some extent, disease specific (Table 1). They include skin prick testing (SPT) with whole allergen extracts, prick-to-prick testing with fresh fruits/vegetables, atopy patch testing and serum specific IgE for whole allergen extracts and component resolved diagnostic testing. Challenges may be undertaken in an appropriate clinical context.

Careful history taking is essential for targeted specific IgE testing. It is recommended that SPT for anaphylaxis, especially to foods and drugs, should be performed with facilities available for resuscitation because severe systemic allergic reactions and rare deaths have occurred (12). Multifactorial anaphylaxis due to co-factors may involve challenge with the suspected triggers concurrently if allergen testing is negative. Testing for specific IgE to omega-5 gliadin in cereal grain may be relevant to patients with a history of exercise-induced anaphylaxis or tropomyosin in patients with house dust mite sensitization.

The relevance of patient and environmental factors in interpreting IgE sensitization test results

Personal, family and medical features of a patient's history are the paramount considerations in determining when to test for allergic sensitization because these factors contribute significantly to the ability of the clinician to correctly interpret these tests.

Personal factors

It is well known from both clinic- and population-based studies that the prevalence of the disease in a population strongly influences the usefulness of the disease-specific tests. Screening of non-symptomatic individuals with specific IgE tests has a low positive predictive value, in the range of 50%. In clinic populations, where the likelihood of the disease being present is higher, the positive predictive value of skin prick testing (SPT) or specific IgE testing may be higher than 85% (13). In contrast, in both unselected and referred populations the tests have high negative predictive value and specificity.

Individuals from different ethnic backgrounds may have different tendencies to develop or demonstrate positive skin test and specific IgE results. This has been shown recently in the screening data from the LEAP study where higher rates of specific IgE were found in black individuals than in other ethnic groups (14). Also specific IgE levels for peanut are higher in African-American subjects than in non-African-American subjects in US studies (15).

It has been extensively documented that a subject's age has a considerable impact on the interpretation of tests, for example, a difference of a year may double the likelihood of clinical allergy in pre-school children (16)

Other patient factors include the presence or absence of associated diseases such as atopic dermatitis/eczema. There is a strong association between eczema severity and food allergy (14, 17); this may relate to underlying filaggrin mutations (18).

Environmental factors

Geographical location of patients may make considerable differences. Important factors are presumably temperature, humidity and pollen exposure. In Australia, rates of peanut allergy are higher in the South than the North (19) and specific IgE testing for peanut shows

different patterns from North and Western to Southern Europe (20,21); Ara h 2 is the dominant allergen in the North and West, but being of less significance in Southern Europe (22). Similarly, the EPAAC study showed wide variations in peanut specific IgE levels across European countries in a pre-school population with moderately severe eczema (23).

Other factors

Potential co-factors also need to be considered when interpreting the clinical history in relation to IgE test results. Exercise in the few hours after eating a specific food to which the individual is sensitized might be necessary in order to trigger a food-related reaction (24). Other cofactors include physical and emotional stress, infections and medications. Co-existing morbidities such as mastocytosis in beekeepers can definitely modify the pattern of reaction and may influence IgE testing interpretation (25).

Finally, IgE testing may also be used for the follow-up of an allergic disease. Changes in results with sequential testing in individual patients can indicate whether food allergies are likely to persist or resolve (26, 27). They can be used to decide if repeat food challenge is needed for assessing if a child has outgrown a specific food (28).

Maximizing the available diagnostic information by treating SPT and specific IgE results as continuous measures

Traditionally, skin prick testing and specific IgE results have been dichotomised into positive or negative according to an arbitrary cut off value. Typically, this was a 3mm wheal diameter for skin prick testing and 0.35 kU/l for specific IgE. This has given rise to the use of sensitivity (proportion of true positives that are correctly identified) and specificity (proportion of true negatives that are correctly identified) as a way of describing the utility of the test

(10). There is a trade-off in terms of how useful tests are, those with good sensitivity usually have poor specificity and visa versa. Sensitivity and specificity provide a population perspective, while this may be useful for public health screening tests, it is less helpful for interpreting test results for individual patients. So a patient with a SPT ≥ 3 mm or specific IgE of ≥ 0.35 kU/L to a food, only has an approximate 50% chance of being allergic; meanwhile another patient with a 0mm SPT or < 0.35 specific IgE result still has approximately a 10% chance of being allergic to that food.

The introduction of 95% positive predictive values (29) has helped clinicians to provide more clinically relevant recommendations from the test results. An example would be 15kU/L for peanut, children with a result at or above this value have a 95% chance of having peanut allergy (30), there are many others examples in the literature (3). This approach provides a patient rather than a population perspective. However, most patients have results that fall into the grey area below the 95% positive predictive value (29) (Figure 2), the confidence intervals associated with these positive predictive values are wide so they are not as precise as they may seem to be (31) and they do not take into account other factors such as presenting history.

In a further refinement of the interpretation of allergy test results, the relationship between the result and the probability of clinical allergy has been modelled using logistic regression. This approach gives curves where the probability of clinical allergy can be estimated for different test results (32, 30) (Figure 3). While this appears to maximise the information available to guide clinical management, these probability curves only relate to the relationship between test result and clinical allergy in an “average” patient. Patients seen in an allergy clinic are very heterogeneous, for example, some may have a non-allergic underlying disorder and others may be sensitized but not clinically allergic.

Many groups have looked to see whether IgE test results can provide information about the likely severity of future allergic reactions. In general, no relationship has been found (32) although one study found a weak relationship with more severe reactions being associated with slightly larger SPT results (33).

Utilizing allergen components to improve our diagnostic approach

Traditional diagnostic tests in allergy are directed towards crude extracts which are made up of a number of different allergenic molecules. These different molecules can be individually assayed using component-resolved diagnostics (CRD) (34). CRD can discriminate between binding to potent, more relevant allergens and cross-reactive, clinically less relevant allergens. In allergen extracts some allergens are present in a sub-optimal concentration. Allergen extracts contain, in addition to the allergens, many other non-allergenic components that might hamper the analysis of IgE binding to the relevant allergens, the CRD approach avoids this issue.

CRD has introduced improvements in allergy diagnostics, especially in the areas of respiratory (35) and food allergies. For instance, more than 10 different allergenic peanut proteins are described: Ara h 1 to Ara h 13. Within this group it seems that IgE binding to Ara h 2 and Ara h 6 allergens, belonging to the conglutin protein family, are the most relevant to clinical peanut allergy (36). The Ara h 8 allergen in peanut belongs to the PR-10 family, like Bet v 1 the major allergen in birch pollen. This can cause false positive test results when patients with birch pollen allergy are tested with whole peanut extract. Additionally, some of the allergens in food can predict severe type of reactions. This has been shown for apple where Mal d 1-specific IgE is associated with mild, local allergic reactions, while Mal d 3-specific IgE is associated with more severe, systemic reactions. Also in hazelnut allergy, Cor a 1 is associated with mild while Cor a 9 and Cor a 14 are associated with more severe allergic reactions (37, 38).

CRD can also be used to define the most relevant allergen for allergy immunotherapy. For instance, in the case of insect venom allergy with clinical non-relevant cross-reactivity between bee and wasp allergens, the determination of IgE to the bee venom protein Api m 1 and wasp venom allergens Ves v 1 and Ves v 5 can be helpful in determining the clinically relevant sensitization (39).

In a diagnostic setting there are a number of ways that CRD can be utilised. The most widely used approach is determination as single component-specific-IgE (e.g. ImmunoCAP, ImmuLite, HyTech) (40). Single determination can also be performed by the basophil activation test, although the latter is less available in routine settings. To fully utilise the potential of CRD, a number of IgE components need to be assayed per patient in a multiplex analysis. This can be done in a multiplex system, currently available as the semi-quantitative ImmunoCAP-ISAC system which enables the determination of IgE binding to 112 allergens in one test run using only 30 microlitres of serum. An important difference between single and multiplex systems is the amount of allergen, the ISAC system uses approximately 100 000 fold less allergen than the ImmunoCAP (40, 41). This may underlie its lower sensitivity at low sIgE levels as there will be more competition between allergen specific IgG and IgE to bind to the lower amount of immobilized allergen on the ISAC chip (40). Also the allergens coupled to the single allergen system are not always the same as on the multiplex system and allergens will differ between manufacturers.

The multiplex analysis approach generates multiple data points per patient, this has important implications for how these data are interpreted. Results for 112 individual allergens might be too complicated to interpret with many clinically irrelevant sensitizations. An approach where the focus is on IgE binding to specific groups of allergens maybe more helpful and diagnostic algorithms are currently being developed to do this.

Potential for IgG to confound the result of sIgE assay systems

The level of specific IgE may be underestimated by the presence of other allergen-specific antibodies, particularly by specific IgG's including IgG4. It is also likely that allergen-blocking antibodies mitigate the clinical effects of specific IgE (42). For this reason, it has been argued that susceptibility to interference makes the outcome of a specific IgE test clinically more relevant. However, such interference by allergen-blocking antibodies is undesirable from the analytical point of view, because the outcome of the assay is not accurately reflecting the level of specific IgE in the sample. With high levels of allergen in the testing system (eg UniCAP system) interference by allergen-blocking antibody is unlikely. Where allergen levels are lower (eg ISAC), the risk of interference increases (43). This may particularly important following specific immunotherapy where specific IgG levels are elevated.

Integrating patient, environmental and allergen factors with IgE test results to predict the likelihood of clinical allergy

Probability curves provide an estimate of the risk of clinical allergy for specific IgE test results (Figure 3) for the “average” patient. The chance of clinical allergy can though be more precisely estimated from a test result by taking into account the presenting features of a patient; this is the so-called pre-test probability (44, 9). A nomogram approach to interpreting test results in relationship to the pre-test probability has been developed (45). This calculates the chance of clinical allergy using a likelihood ratio which is the likelihood of a given test result in clinically allergic subject over the likelihood of a similar result in normal subject (see example in Figure 4). Although this approach maximised the available clinical information, it has not been widely adopted by clinicians, perhaps because it is too time consuming, too complicated or clinicians do not have good estimates for pre-test probabilities. There are likelihood ratio data for a limited number of foods (46) and almost no

Accepted Article

data for aeroallergens. A problem with many of the studies in which likelihood ratios (or other diagnostic parameters) are calculated is that subjects with very different pre-test probabilities are mixed (47). Additionally we do not know if pre-test probabilities are stable across different populations.

An alternative approach would be to use modern computer power to generate programmable calculator systems to synthesise presenting features together with the allergy test results to give a prognostic indication of the likelihood of clinical allergy. These are underpinned by logistic regression models built from clinical cases. An example is the Cork / Southampton prognostic model for food allergy (48). A proto algorithm was generated using data from Southampton food challenges (n=429, peanut, egg, milk). It was then retrospectively applied, evaluated and modified using data from Cork food challenges (n=289, peanut, egg, milk). Finally it was prospectively validated in a blinded study in a further group of Cork children undergoing food challenges (n=70, peanut, egg, milk). The clinical features that were found to predict clinical allergy were symptoms, sex, age, skin prick test, specific IgE and total IgE. The model had 97% accuracy for allergy and 94% for tolerance with an area under the curve of 0.97 for peanut, 0.95 for egg and 0.94 for milk in the prospective validation study. The model has since been validated in other settings (49).

The major drawback of the both the nomogram and calculator approach is the need to be able to derive a pre-test probability for individual patients. While these have been developed for specific settings (48), we do not know how they differ between different setting and regions (49). Without a good estimate of the pre-test probability, the predictive power of the system will be poor. Further data from a diverse range of setting, regions and allergens is required before this approach can be used routinely. It does though show promise.

IgE sensitization test results from the patient perspective

Patients generally regard IgE sensitization tests as allergy tests and they directly self-translate results into clinical diagnoses. To help patients understand the meaning of their IgE sensitization test results, clinicians need to explain the nature of these tests before ordering them and not refer to them as allergy tests. Such a clear understanding is essential if patients are going to have the confidence to implement changes to their self-management plans. On the face of it, a negative IgE sensitization test result might be expected to exclude clinical allergy. Patients need to understand that these tests can not completely exclude allergy, especially when their clinical history is very suggestive of a clinical allergy. This situation needs to be managed with the patient, for example would it be safe to re-introduce the food at home in a graded manner or should a food challenge be ordered? Patients need to understand that sometimes tests give inconclusive results that do not allow clinicians to rule out or rule in clinical allergy. This situation would warrant a discussion about the merits of a provocation challenge to establish a firm conclusion. Finally, even where the IgE sensitization test is positive, a dialogue with the patient is needed as not all will have results that are overwhelming predictive of clinical allergy. Is the level of certainty sufficient for an individual patient to be happy to avoid the allergen or is a provocation challenge required to firmly establish the diagnosis? Individual patients may have different preferences about which IgE sensitization test is used and how they are subsequently managed in the context of different test results.

Potential new advances in allergy diagnostics

Novel approaches to the diagnosis of IgE-mediated allergic disease have been researched in the recent years, including the basophil activation test (BAT), IgE and IgG4 binding to linear allergen peptides, mediator release and genetic profiling.

BAT is a flow cytometry-based assay that assesses changes in the expression of activation markers on the surface of basophils following allergen-stimulation (51). Being a functional assay, it has the potential to more closely resemble the clinical phenotype of patients than methods that merely detect the presence of IgE. BAT has been applied to the diagnosis of allergy to food as well as drug, venom and environmental allergens (52). Its utility seems to be best in cases where conventional allergy tests have failed to diagnose allergy and provocation tests are to be considered (53). BAT could allow a reduction in the need for provocation tests (53). It seems to be valuable also in monitoring patients undergoing immuno-modulatory treatments such as allergen-specific immunotherapy (54, 55) and omalizumab (56), and in monitoring natural resolution of food allergy (57, 58).

IgE and IgG4 binding to linear peptides has been studied in food and respiratory allergies. IgG4 do not seem to have a role in diagnosing allergies (59). Allergic patients tend to have greater IgE binding and broader epitope diversity than tolerant patients (60). Higher epitope diversity has also been associated with more severe reactions to food (61). For different allergens, immunodominant epitopes have been identified. Technical challenges as well as cost-effectiveness, role of tertiary structures and need for validation have hampered its wider use in clinical practice (47).

Other methods such as mediator levels, such as PAF (62, 63), and genetic profiling (64) may be useful in predicting patients at risk of developing more severe allergic reactions in the future.

Summary and future perspectives

Over the last decade there have been considerable advances in our understanding of the immunological changes underlying clinical allergy and the relationship between skin prick and specific IgE test results and clinical manifestations of allergy. We have therefore re-

considered our approach to interpreting specific IgE test results and how we utilize them in clinical practice (Box 2). The meaning of a specific IgE sensitization test results varies according to the prevalence of allergy in the test population, a patients' age, their ethnicity and any co-existing clinical diseases such as eczema (Figure 5). The history may also indicate that a co-factor must be present for a clinical reaction to occur when a patient with specific IgE to the allergen to which they are exposed. We now have data to map specific IgE sensitization test results to the probability of clinical allergy for several foods. Unfortunately, these do not take into account patient specific factors, neither do the 95% positive predictive values that are routinely used in clinical practice.

The chance of clinical allergy can be more precisely estimated from an IgE sensitization test result, by taking into account the presenting features of a patient (pre-test probability). The presence of each of these patient specific factors may mean that a patient is more or less likely to have clinically allergy with a given test result (Figure 5). Together they may considerably modify the interpretation of IgE sensitization test results. We have presented two approaches to including pre-test probabilities in the interpretation of results. The nomogram approach employs likelihood ratios for specific test results to translate pre-test probabilities into the chance of clinical allergy (post-test probability). This approach has not been widely implemented, perhaps because of the lack of likelihood ratio data and its apparent complexity. Computer based calculator systems can overcome these issues but they rely on internal models to generate pre-test probabilities from similar patient information, these are limited by a lack of data from a wide range of diverse setting, regions and allergens. We need more studies to determine to what extent the meaning of test results differ in different populations. Finally a simpler approach has been proposed where each of the presenting history and IgE sensitization test results are divided into three levels of likelihood for clinical allergy and these are synthesised in a 3 by 3 table (9). This approach has not though been formally validated.

Accepted Article

It is very apparent that there are gaps in our knowledge about the role of IgE in allergic disease and our interpretation of IgE sensitization tests. In IgE-mediated allergy, IgE is necessary but not sufficient to elicit an allergic reaction. A better understanding of the mechanisms of IgE-mediated allergic disease would help us to interpret false-positive IgE levels. This includes tissue factors that may play a role at the effector cell level but also patient factors (e.g. ethnicity, co-existing inflammatory diseases) and environmental factors (e.g. migration, climate change, cultural characteristics) that can function as a promoter or an enhancer of the allergic response.

Component resolved diagnosis (CRD) represents an important advance in specific IgE testing. Clinically the ability to be able to discriminate between binding to potent, more relevant allergens and cross-reactive, clinical less relevant allergens is exciting. The implementation of the technology is currently limited by the lack of studies that focus on whether the CRD approach can improve the diagnostic experience of patients. We also await the availability of commercial CRD SPT reagents. For multiplex CRD systems we also need to develop better approaches to interpreting IgE sensitization data on over 100 allergens to avoid over diagnosing clinical allergy.

Lastly we need better systems and approaches to identify clinical important IgE sensitization. The BAT and IgE binding to linear allergen peptides are new diagnostic tests currently in development. Many patients have symptoms of IgE-mediated allergy without evidence of specific IgE sensitization. This may be due to the absence of the allergens that are driving the symptoms in the diagnostic extracts or to local production of allergen-specific IgE in the target organ. The utility of tests other than IgE sensitization tests, such as the atopy patch test, in the diagnosis of conditions with mixed IgE and non-IgE-mediated pathogenesis, such as atopic dermatitis and eosinophilic esophagitis, should also be explored further.

Acknowledgements

We would like to acknowledge the support of EAACI in developing this position paper.

Author contributions

This paper was generated by a pan-EAACI task force supported by the Asthma Section, Dermatology Section, ENT Section, Immunology Section, Pediatric Section, Allergy Diagnosis Interest Group, Allied Health Interest Group, Drug Allergy Interest Group, Food Allergy Interest Group, JMA Working Group and Patient Group. The task force chair was Graham Roberts and the secretary was Markus Ollert. Each author drafted a section which Graham Roberts edited into the final document. All authors reviewed and discussed the final document and approved the final version. The final version was evaluated and endorsed by the EAACI Executive Committee.

Conflicts of interest

Maira Austin has received support from ALK-Abello to attend BSACI meeting. Philippe Eigenmann has received lecture honorarium and research grants from ThermoFisher Scientific and is on the scientific advisory board of MicrotestDX. Edward F. Knol has received speakers fee and travel reimbursement from Thermo Fisher Scientific. Graham Roberts has received funding for his research programme and acted as a scientific consultant for ALK-Abello and Thermo Fisher. Alexandra F. Santos has received research funding from the Medical Research Council, UK; the NIAID and the Immune Tolerance Network, USA; and from the National Peanut Board, USA. She has received research support for a multicentre project using basophil activation test from Bühlmann Laboratories

AG, Switzerland, through King's College London. Rob Aalberse has no conflict of interest to declare.

References

1. Papadopoulos NG, Agache I, Bavbek S, Bilo BM, Braidó F, Cardona V, et al. Research needs in allergy: an EAACI position paper, in collaboration with EFA. *Clin Transl Allergy*. 2012; 2: 21.
2. Johansson SG, Bieber T, Dahl R, Friedmann PS, Lanier BQ, Lockey RF, et al. Revised nomenclature for allergy for global use: Report of the Nomenclature Review Committee of the World Allergy Organization, October 2003. *J Allergy Clin Immunol* 2004; 113: 832-6.
3. Bousquet J, Heinzerling L, Bachert C, Papadopoulos NG, Bousquet PJ, Burney PG, et al. Practical guide to skin prick tests in allergy to aeroallergens. *Allergy* 2012; 67: 18–24.
4. Stiefel G, Roberts G. How to use serum specific IgE measurements in diagnosing and monitoring food allergy. *Arch Dis Child Educ Pract Ed* 2012; 97: 29–36.
5. Werfel T., Ballmer-Weber B., Eigenmann P. A., Niggemann B., Rancé F., Turjanmaa K. and Worm M. Eczematous reactions to food in atopic eczema: position paper of the EAACI and GA2LEN. *Allergy* 2007; 62: 723–728.
6. Patil SU, Shreffler WG. BATting above average: Basophil activation testing for peanut allergy. *J Allergy Clin Immunol* 2014;134:653-4.
7. Eigenmann PA, Atanaskovic-Markovic M, O'B Hourihane J, Lack G, Lau S, Matricardi PM, et al. Testing children for allergies: why, how, who and when: An updated statement of the European Academy of Allergy and Clinical Immunology (EAACI) Section on Pediatrics and the EAACI-Clemens von Pirquet Foundation. *Pediatr Allergy Immunol* 2013; 24:195-209.

8. Scadding G, Hellings P, Alobid I, Bachert C, Fokkens W, van Wijk R et al. Diagnostic tools in Rhinology, EAACI position paper. Clin Transl Allergy 2011;1:2.
9. Muraro A*, Agache I*, Clark A*, Sheikh A, Roberts G, Akdis CA, et al. Management patients with food allergy in the community. Allergy 2014; 69: 1046–1057.
10. Soares-Weiser K, Takwoingi Y, Panesar SS, Muraro A, Werfel T, Hoffmann-Sommergruber K, et al. The diagnosis of food allergy: a systematic review and meta-analysis. Allergy 2014; 69: 76-86.
11. Fujita H, Soyka MB, Akdis M, Akdis C. Mechanisms of allergen-specific immunotherapy. Clin Transl Allergy 2012; 2:2.
12. Heinzerling L, Mari A, Bergman K-C et al. The skin prick test – European Standards. Clin Transl Allergy 2013; 3:3.
13. Miceli Sopo S, Radzik D, Calvani M. The predictive value of specific immunoglobulin E levels for the first diagnosis of cow's milk allergy. A critical analysis of pediatric literature. Pediatr Allergy Immunol 2007; 18: 575–582.
14. Dutoit G*, Roberts G*, Sayre PH, Plaut M, Bahnson T, Mitchell H, et al. Identifying infants at high risk of peanut allergy – The LEAP Screening Study. J Allergy Clin Immunol 2013; 131: 135-43.e1-12.
15. Branum AM, Lukacs SL. Food Allergy Among Children in the United States. Pediatrics 2009;124:1549-1555
16. Komata T. Soderstrom L, Borres MP, Tachimoto H, Ebisawa M. The predictive relationship of food-specific serum IgE concentrations to challenge outcomes for egg and milk varies by patient age. J Allergy Clin Immunol 2007; 119: 1272-4.

- Accepted Article
17. van Veen WJ, Dikkeschei LD, Roberts G, Brand PLP. Predictive value of specific IgE for clinical peanut allergy in children: relationship with eczema, asthma, and setting (primary or secondary care). *Cli Translational Allergy* 2013; 3: 34.
 18. Irvine AD, Irwin McLean W.H, Leung DY.M. Filaggrin Mutations Associated with Skin and Allergic Diseases. *N Engl J Med* 2011; 365:1315-1327.
 19. Osborne NJ, Ukoumunne OC, Wake M, Allen KJ. Prevalence of eczema and food allergy is associated with latitude in Australia. *J Allergy Clinical Immunol* 2012; 129: 865–867.
 20. Nwaru BI, Hickstein L, Panesar SS, Muraro A, Werfel T, Cardona V, et al. The epidemiology of food allergy in Europe: a systematic review and meta-analysis. *Allergy* 2014; 69: 62-75.
 21. Nwaru BI, Hickstein L, Panesar SS, Roberts G, Muraro A, Sheikh A on behalf of The EAACI Food Allergy & Anaphylaxis Guidelines Group. Prevalence of common food allergies in Europe: a systematic review and meta-analysis. *Allergy*, *in press*.
 22. Hourihane JO'B. Peanut allergy. *Pediatric Clinics of North America* 2011; 58:445-458.
 23. de Benedictis FM, Franceschini F, Hill D, Naspitz C, Simons FE, Wahn U, Warner JO, et al. The allergic sensitization in infants with atopic eczema from different countries. *Allergy* 2009; 64: 295-303.
 24. Du Toit G. Food-dependent exercise-induced anaphylaxis in childhood. *Pediatr Allergy Immunol* 2007;18: 455-63.
 25. Bonadonna P, Zanotti R, Müller U. Mastocytosis and insect venom allergy. *Curr Opin Allergy Clin Immunol* 2010; 10: 347-53.
 26. Ho MH, Wong WH, Heine RG, Hosking CS, Hill DJ, Allen KJ. Early clinical predictors of remission of peanut allergy in children. *J Allergy Clin Immunol* 2008;121:731–6.

27. Skripak JM1, Matsui EC, Mudd K, Wood RA. The natural history of IgE-mediated cow's milk allergy. *J Allergy Clin Immunol* 2007; 120: 1172-7.
28. Shek LP, Soderstrom L, Ahlstedt S, Beyer K, Sampson HA. Determination of food specific IgE levels over time can predict the development of tolerance in cow's milk and hen's egg allergy. *J Allergy Clin Immunol* 2004; 114: 387-91
29. Roberts G, Lack G. Food allergy - getting more out of your skin prick tests. *Clin Exp Allergy* 2000; 30: 1495-8.
30. Sampson HA. Utility of food-specific IgE concentrations in predicting symptomatic food allergy. *J Allergy Clin Immunol*. 2001; 107: 891-6.
31. Roberts G, Lack G. Diagnosing peanut allergy with skin prick and specific IgE testing. *J Allergy Clin Immunol* 2005;115:1291-6.
32. Sampson HA, Ho DG. Relationship between food-specific IgE concentrations and the risk of positive food challenges in children and adolescents. *J Allergy Clin Immunol* 1997; 100: 444-51.
33. Houriahane JO'B, Kilburn SA, Dean P, Warner JO. Clinical characteristics of peanut allergy. *Clinical & Experimental Allergy* 1997; 27: 634–639.
34. Valenta R, Lidholm J, Niederberger V, Hayek B, Kraft D, Grönlund H. The recombinant allergen-based concept of component-resolved diagnostics and immunotherapy (CRD and CRIT). *Clin Exp Allergy* 1999; 29: 896-904.
35. Douladiris N, Savvatanos S, Roumpedaki I, Skevaki C, Mitsias D, Papadopoulos N.G. A Molecular Diagnostic Algorithm to Guide Pollen Immunotherapy in Southern Europe: Towards Component-Resolved Management of Allergic Diseases. *Int Arch Allergy Immunol* 2013; 162: 163-172

- Accepted Article
36. Klemans RJ, Knol EF, Bruijnzeel-Koomen CA, Knulst AC. The diagnostic accuracy of specific IgE to Ara h 6 in adults is as good as Ara h 2. *Allergy* 2014; 69: 1112-4.
37. Fernández-Rivas M, Bolhaar S, González-Mancebo E, Asero R, van Leeuwen A, Bohle B, et al. Apple allergy across Europe: how allergen sensitization profiles determine the clinical expression of allergies to plant foods. *J Allergy Clin Immunol* 2006; 118: 481-8.
38. Masthoff LJ, Mattsson L, Zuidmeer-Jongejan L, Lidholm J, Andersson K, Akkerdaas JH, et al. Sensitization to Cor a 9 and Cor a 14 is highly specific for a hazelnut allergy with objective symptoms in Dutch children and adults. *J Allergy Clin Immunol* 2013; 132: 393-9.
39. Müller U, Schmid-Grendelmeier P, Hausmann O, Helbling A. IgE to recombinant allergens Api m 1, Ves v 1, and Ves v 5 distinguish double sensitization from crossreaction in venom allergy. *Allergy* 2012; 67: 1069-73.
40. Canonica GW, Ansotegui IJ, Pawankar R, Schmid-Grendelmeier P, van HM, Baena-Cagnani CE et al. A WAO - ARIA - GA²LEN consensus document on molecular-based allergy diagnostics. *A. World Allergy Organ J* 2013; 6:17.
41. Melioli G, Bonifazi F, Bonini S, Maggi E, Mussap M, Passalacqua G et al. The ImmunoCAP ISAC molecular allergology approach in adult multi-sensitized Italian patients with respiratory symptoms. *Clin Biochem* 2011; 44: 1005-11.
42. Geroldinger-Simic M, Zelniker T, Aberer W, Ebner C, Egger C, Greiderer A, et al. Birch pollen-related food allergy: clinical aspects and the role of allergen-specific IgE and IgG4 antibodies. *J Allergy Clin Immunol*. 2011;127:616-22
43. Lupinek C, Wollmann E, Baar A, Banerjee S, Breiteneder H, Broecker BM, et al. Advances in allergen-microarray technology for diagnosis and monitoring of allergy: The MeDALL allergen-chip. *Methods* 2013; 66: 106-119.

- Accepted Article
44. Du Toit G, Santos A, Roberts G, Smith P, Fox AT, Lack G. The diagnosis of IgE-mediated food allergy in childhood. *Pediatric Allergy Immunol* 2009; 20:309-19.
 45. Fagan TJ. Letter: Nomogram for Bayes theorem. *N Engl J*
 46. Peters RL, Gurrin LC, Allen KJ. The predictive value of skin prick testing for challenge-proven food allergy: A systematic review. *Pediatr Allergy Immunol* 2012; 23: 347–352.
 47. Aalberse RC. Food-derived peptides in a diagnostic context: the fewer the better? *Pediatr Allergy Immunol*. 2014;25:206-7).
 48. DunnGalvin A, Daly D, Cullinane C, Stenke E, Keeton D, Erlewyn- Lajeunesse M, Roberts GC, Lucas JSA, Hourihane J'OB. Highly accurate prediction of food challenge outcome using routinely available clinical data. *J Allergy Clin Immunol* 2011; 127, 633-639.
 49. Klemans RJB, Otte D, Knol M, Knol EF, Meijer Y, Gmelig-Meyling FHJ. The diagnostic value of specific IgE to Ara h 2 to predict peanut allergy in children is comparable to a validated and updated diagnostic prediction model *J Allergy Clin Immunol* 2013; 131: 157-63
 50. Roberts G. Anaphylaxis to foods. *Pediatr Allergy Immunol* 2007; 18: 543–548. *Med* 1975;293:257.
 51. Ebo DG, Bridts CH, Hagendorens MM, Aerts NE, De Clerck LS, Stevens WJ. Basophil activation test by flow cytometry: present and future applications in allergology. *Cytometry B Clin Cytom* 2008; 74:201-10.
 52. Hoffmann HJ, Santos AF, Mayorga C, et al. Clinical impact of the basophil activation test in the the diagnosis and monitoring of allergic disease. *Allergy* (in preparation)
 53. Santos AF, Douiri A, Becares N, Wu S-Y, Stephens A, Radulovic S, et al. Basophil activation test discriminates between allergy and tolerance in peanut-sensitized children. *J Allergy Clin Immunol* 2014; 134: 645-52.

54. Jones SM, Pons L, Roberts JL, Scurlock AM, Perry TT, Kulis M, et al. Clinical efficacy and immune regulation with peanut oral immunotherapy. *J Allergy Clin Immunol* 2009; 124:292-300, e1-97.
55. Thyagarajan A, Jones SM, Calatroni A, Pons L, Kulis M, Woo CS, et al. Evidence of pathway-specific basophil anergy induced by peanut oral immunotherapy in peanut-allergic children. *Clin Exp Allergy* 2012; 42:1197-205.
56. Nopp A, Johansson SGO, Ankerst J, Bylin G, Cardell LO, Grönneberg R et al. Basophil allergen threshold sensitivity. A useful approach to anti-IgE treatment efficacy evaluation. *Allergy* 2006;61:298–302.
57. Rubio A, Vivinus-Nebot M, Bourrier T, Saggio B, Albertini M, Bernard A. Benefit of the basophil activation test in deciding when to reintroduce cow's milk in allergic children. *Allergy* 2011; 66:92-100.
58. Wanich N, Nowak-Wegrzyn A, Sampson HA, Shreffler WG. Allergen-specific basophil suppression associated with clinical tolerance in patients with milk allergy. *J Allergy Clin Immunol* 2009; 123:789-94 e20.
59. 17. Muraro A*, Werfel T*, Roberts G, Beyer K, Bindslev Jensen C, Cardona V, et al on behalf of EAACI Food Allergy and Anaphylaxis Guidelines Group. Diagnosis and management of food allergy: Guidelines from the European Academy of Allergy and Clinical Immunology. *Allergy*, 2014; 69: 1008-1025.
60. Lin J, Bruni FM, Fu Z, Maloney J, Bardina L, Boner AL, et al. A bioinformatics approach to identify patients with symptomatic peanut allergy using peptide microarray immunoassay. *J Allergy Clin Immunol* 2012; 129:1321-8 e5.

61. Flinterman AE, Knol EF, Lencer DA, Bardina L, den Hartog Jager CF, Lin J, et al. Peanut epitopes for IgE and IgG4 in peanut-sensitized children in relation to severity of peanut allergy. *J Allergy Clin Immunol* 2008; 121:737-43 e10.
62. Vadas P, Gold M, Perelman B, Liss GM, Lack G, Blyth T, et al. Platelet-activating factor, PAF acetylhydrolase, and severe anaphylaxis. *N Engl J Med* 2008; 358:28-35.
63. Vadas P, Perelman B, Liss G. Platelet-activating factor, histamine, and tryptase levels in human anaphylaxis. *J Allergy Clin Immunol* 2013; 131, 144–149.
64. Teodosio C, García-Montero AC, Jara-Acevedo M, et al. Gene expression profile of highly purified bone marrow mast cells in systemic mastocytosis. *J Allergy Clin Immunol* 2014; 131: 1213-1224.

Figure 1. Approaches to IgE sensitization testing in a patient with possible allergy

If multiple testing is undertaken without an allergy-focused history, tests are more likely to give false positive results impeding the clinical management of the patient. If an allergy-focused history is used to choose relevant tests, results are more likely to be clinically relevant.

Figure 2. Distribution of skin prick test (SPT) results in a group of 100 hypothetical clinic patients investigated for peanut allergy

Black: SPT \geq 8mm; grey: SPT 3-7mm, white: SPT < 3mm (reproduced from Roberts 2000 with permission (29)).

Figure 3. Probability of food allergy at different specific IgE values.

Reproduced from Sampson 2001 with permission (30).

Figure 4. Illustration of the use of pre-test probabilities and likelihood ratios to diagnosis clinical allergy. (50)

A 6 year old boy develops an urticarial rash about an hour after consuming a little of a peanut snack bar. On the basis of history alone, there is approximately a 25% chance that he is clinically allergic to peanut. He has a 4mm wheal to peanut which has a likelihood ratio for clinical allergy of 2.4 (31). From the nomogram, it can be seen that the probability that he is clinically allergic to peanuts is around 50%. A peanut challenge would be required to make a firm diagnosis. However, if the same child had suffered another similar allergic reaction a few weeks later on contact with peanuts, the pre-test probability would increase to around 80%. With a similar skin prick test result the post-test probability would go up to 90%, a reasonably high level of certainty meaning that a challenge would not be indicated.

Figure 5. Conceptual figure to illustrate the proposed approach

① The nature of the clinical history should determine the best IgE sensitization test. ② Each IgE sensitization test result is associated with a specific probability of clinical allergy as shown by the interrupted red arrows. ③ The exact relationship between test result and probability of allergy varies according to internal and external factors which may make clinical allergy more or less likely in a potentially additive fashion. ④ So the likelihood of clinical allergy can be determine from the IgE sensitization test result and a knowledge of other important patient factors; these precise relationships still need to be established.

Table 1. Investigations to consider for different presentations of IgE disease (7)

Disease	Skin prick tests (SPT)	Intradermal tests (IDT)	Atopy patch test (ATP)	Serum specific IgE	Challenge
Acute urticaria	(+)	-	-	(+)	(+)
Histaminergic angioedema (without wheals)	(+)	-	-	(+)	(+)
Atopic dermatitis	+	-	+	+	+
IgE-mediated food allergies	+	-	-	+	+
Oral allergy syndrome	+ ¹	-	-	+	(+)
Eosinophilic oesophagitis	+	-	(+)	+	-
Asthma	+	-	-	+	(+)
Rhinitis	+	-	-	+	(+)
Conjunctivitis	+	-	-	+	(+)
Anaphylaxis	+	(+)	-	+	(+)
Venom allergy	+	+	-	+	(+)
Drug reactions	+/-	+/-	-	+/-	(+)

Symbols: +: indicated; (+) consider when symptoms are likely to have been precipitated by an allergen or other tests have not provided a clear allergic diagnosis; -: not indicated. ¹ Skin prick tests using fresh fruits or vegetables require a prick-to-prick technique.

Box 1. Examples of organ-specific effects of immediate hypersensitivity reactions

Skin

The relevance of specific IgE sensitization is often unclear in patients with *atopic dermatitis* without a history of associated urticaria closely related to allergen exposure as patients often have multiple sensitizations to inhalant and food allergens. Delayed eczematous reactions up to 24 hours after exposure to relevant allergens, such as house dust mite, grass pollen or cows' milk, are thought to be due to T-cell activation through allergen binding to specific IgE on FcεRI of inflammatory dendritic epidermal cells (IDEC) or Langerhans' cells by antigen focusing. Atopy patch testing maybe used but this has low sensitivity although specificity is reasonable.

Although some *acute urticaria* is caused by immediate hypersensitivity reactions to foods or drugs, the majority of cases appear to relate to acute viral infection (usually of the upper respiratory tract) or remain idiopathic. *Chronic urticaria*, by contrast, is almost never the result of specific IgE sensitization even though activation of FcεRI appears to be a key pathogenic event in some patients with functional autoantibodies directed against FcεRIα or cytophilic IgE. Immunological *contact urticaria* due to environmental exposures or body fluids for example, is probably common but underreported in the community and must be distinguished from non-immunological urticaria that appears to be mast cell and IgE-independent.

Airways

Acute symptoms of allergic *asthma* and *rhinitis* develop after exposure to a number of triggers. One trigger is inhalant allergens to which the patient has developed IgE sensitization. In the nose, this leads to immediate itching, sneezing, rhinorrhoea and congestion whereas the lower airways respond with narrowing, oedema and mucus secretion. Chronic changes result from the influx of inflammatory cells, including eosinophils and damage to epithelial cells. Upper and lower airway symptoms and / or exacerbations

can also occur with other triggers, for example viral respiratory tract infections, cold air or pollution.

Digestive tract

The *oral allergy syndrome* is characterised by immediate oral mucosal symptoms of itching, burning and swelling of the mouth after eating fresh fruits, nuts or vegetables containing pan-allergens (profilins, pathogenesis related protein-10) that are also present in tree or grass pollens to which patients with allergic rhinitis/conjunctivitis have become sensitized. Skin prick testing and serum specific IgE assays may be negative due to loss of allergenicity in standardized reagents. However, prick-to-prick testing with fresh fruits or vegetables will be positive.

Immediate hypersensitivity (*food allergy*) reactions in the digestive tract present acutely with vomiting, diarrhoea and/or abdominal pain. A chronic pattern of inflammation analogous to allergen-induced atopic eczema exacerbations with eosinophilic inflammation has been linked with IgE sensitization, especially in *eosinophilic oesophagitis*, but defining sensitizations by skin prick and specific IgE testing is inconsistently predictive of a response to specific dietary restrictions. Atopy patch testing is often negative.

Systemic reactions

Anaphylaxis is an acute severe systemic reaction involving the skin, airways, bowel and circulation that often presents with syncope or difficulty with breathing. It is usually caused by an allergy to a food, drug or venom. Similar presentations can result from non-allergic mechanisms, with iodinated contrast material for example.

Box 2. Summary

Relevance of IgE sensitization to clinical allergy

- Immediate hypersensitivity (type I) reactions commence with cross-linking of IgE receptors by allergen.
- Specific symptoms of clinical allergy depend on the organ affected.
- Diseases other than allergy may give rise to allergy type symptoms.

Relevance of patient and environmental factors in interpreting IgE sensitization test results

- IgE sensitization tests perform better in clinic populations than the community as the prevalence of symptomatic allergy in a clinic population is higher.
- The meaning of specific IgE sensitization test results varies by patients' age, ethnicity, presenting features of the putative allergic reaction and co-existing clinical diseases such as eczema.
- The presence of co-factors, such as exercise, may be necessary for contact with an allergen to result in an allergic reaction in specific IgE positive patients.

Maximizing the available diagnostic information by treating SPT and specific IgE results as continuous measures

- 95% positive predictive values provide a patient perspective but most patients' results are below these levels and they do not take into account other factors such as the presenting history.
- Probability curves allow the chance of clinical allergy to be estimated from different test results but they do not take into account other factors such as the presenting history.

Utilizing allergen components to improve our diagnostic approach

- Component resolved diagnosis (CRD) can discriminate between binding to potent, more relevant allergens and cross-reactive, clinically less relevant allergens.
- Multiplex CRD systems can generate results for over a hundred component allergens, specific diagnostic algorithms are required to focus attention on the clinically relevant sensitization.
- As current allergen panels are still inadequate, both in number and in biosimilarity,

we still depend on allergen extracts for IgE sensitization testing.

Potential for IgG to confound the result of sIgE assay systems

- The level of specific IgE may be underestimated in the presence of high levels of specific IgG, particularly in assay systems using low amounts of allergen.

Integrating patient, environmental and allergen factors with IgE test results to predict the likelihood of clinical allergy

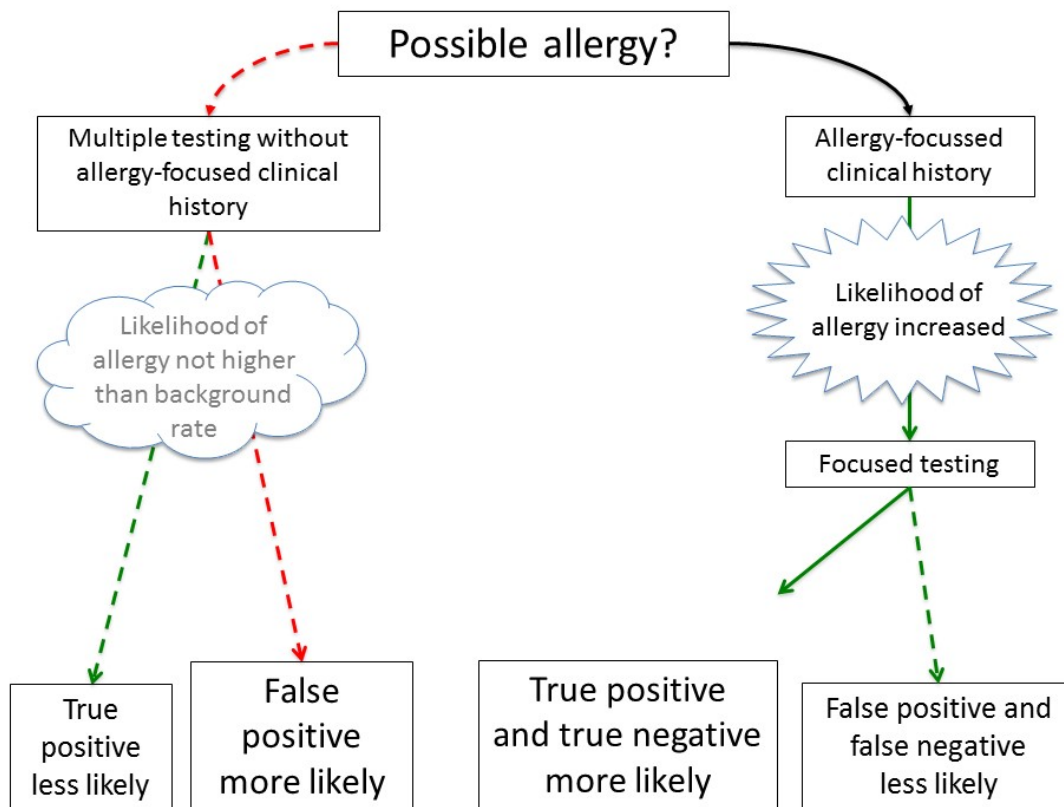
- The chance of clinical allergy can be more precisely estimated from a test result, by taking into account the presenting features of a patient (pre-test probability).
- A nomogram can be used which employs likelihood ratios for specific test results to translate pre-test probabilities to the chance of clinical allergy (post-test probability).
- Computer based calculator systems have been developed to calculate the chance of clinical allergy based on the presenting features of a patient and their IgE sensitization test result.
- This approach is currently limited by a lack of data to allow us to derive pre-test probabilities for diverse setting, regions and allergens.

IgE sensitization test results from the patient perspective

- Patients generally see IgE sensitization tests as being synonymous with diagnostic allergy tests.
- The interpretation of the IgE sensitization test result should be explained to the patient and then there needs to be a two way discussion about subsequent clinical management.

Potential new advances in allergy diagnostics

- Basophil activation test (BAT) may potentially better reflects the presence of clinical allergy than other IgE sensitization tests but further validation is required.
- Other approaches, such as binding to linear peptides and platelet activating factor, still require further developmental work before they can be assessed for their role in the clinic.



SPT ≥ 8 mm = "diagnostic" of peanut allergy

